# Atriopeptin II-induced relaxation of rabbit aorta is potentiated by M&B 22,948 but not blocked by haemoglobin

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- 1 We examined the effects of haemoglobin (which inhibits the vascular responses to stimulation of soluble guanylate cyclase) and of M&B 22,948 (which selectively inhibits cyclic GMP phosphodiesterase) on the relaxation induced in rabbit aorta by the atrial natriuretic peptide, atriopeptin II (which stimulates particulate guanylate cyclase).
- 2 Pretreatment with M&B 22,948 (100 µM) produced a 2.3 fold potentiation of atriopeptin II-induced relaxation of endothelium-denuded rings of rabbit aorta.
- 3 Pretreatment with haemoglobin (10 µM) had no effect on the relaxation or the 10.9 fold increase in cyclic GMP content induced by atriopeptin II in endothelium-denuded rings of rabbit aorta.
- 4 The potentiation by M&B 22,948 suggests a causal role for cyclic GMP in mediating atriopeptin II-induced vasodilatation of rabbit aorta.
- 5 The inability of haemoglobin to block the atriopeptin II-induced rise in cyclic GMP suggests that it does not block stimulation of particulate guanylate cyclase. Thus, it is unlikely that a ferrous haem-containing receptor site is involved in the activation of the particulate form of guanylate cyclase as it is with soluble guanylate cyclase.

## Introduction

Smooth muscle relaxation induced by the endothelium-derived relaxing factor (EDRF) (Furchgott & Zawadzki, 1980), the bovine retractor penis inhibitory factor (IF) (Ambache et al., 1975; Gillespie & Martin, 1980), ultraviolet light (Furchgott et al., 1961), and the nitrovasodilators is associated with an increase in cyclic GMP content (Schultz et al., 1977; Rapoport & Murad, 1983; Bowman & Drummond, 1984; Karlsson et al., 1984; Furchgott et al., 1984b). For the nitrovasodilators, acting via nitric oxide (Arnold et al., 1977; Katsuki et al., 1977), and for EDRF and ultraviolet light, these responses are mediated through activation of the soluble form of the enzyme guanylate cyclase (Rapoport et al., 1983; Karlsson et al., 1984; Busse et al., 1985), though the ability of IF to activate this form of the enzyme has not yet been tested. Soluble guanylate cyclase has a ferrous haem-containing receptor site with which stimulants interact to induce enzyme activation (Craven & De Rubertis, 1978). Exogenously added haemoglobin inhibits the smooth muscle relaxation and concomitant increases in guanosine 3':5'-cyclic monophosphate (cyclic GMP) induced by EDRF, the nitrovasodilators, IF and ultraviolet light (Bowman & Gillespie, 1982; Bowman & Drummond, 1984; Martin et al., 1985, Furchgott et al., 1984b). It has been proposed that this inhibition results from haemoglobin competing with the haem moiety of guanylate cyclase for the active principles (Martin & Smith, 1985; Martin et al., 1986b).

Another class of substances, the atrial natriuretic peptides (ANPs) which appear to be involved in fluid volume regulation (Currie et al., 1983), likewise induce smooth muscle relaxation in association with a rise in cyclic GMP (Fiscus et al., 1985). ANPs do not activate soluble guanylate cyclase, but a particulate form of this enzyme (Waldman et al., 1984; Winquist et al., 1984; Mittal, 1985).

It is not known if particulate guanylate cyclase, like the soluble isoenzyme, has a ferrous haem-containing receptor site, although some indirect evidence against this has been obtained (Rapoport et al., 1985). We investigated this possibility by examining the effects of haemoglobin on vascular relaxation and increases in cyclic GMP content induced by the ANP, atriopeptin II, in endothelium-denuded rings of rabbit aorta.

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## Methods

# Organ bath studies

The preparation of aortic rings was similar to that described by Furchgott & Zawadzki (1980). Briefly, male New Zealand white rabbits weighing 2 to 3 kg were killed by stunning and exsanguination. The aorta was removed, cleaned of adhering fat and connective tissue and cut into 2.5 mm wide transverse rings with a razor blade slicing device. Endothelial cells were removed by gently rubbing the intimal surface with a wooden stick for 30 to 60 s. Successful removal of endothelial cells was confirmed later by the inability of acetylcholine (1 µM) to induce relaxation. Rings were then mounted under 2 g resting tension on stainless steel hooks in 20 ml organ baths, and bathed at 37°C in Krebs solution containing (mm): NaCl 118, KCl 4.8, CaCl<sub>2</sub> 2.5, MgSO<sub>4</sub> 1.2, KH<sub>2</sub>PO<sub>4</sub> 1.2, NaHCO<sub>3</sub> 24, glucose 11 and disodium EDTA 0.03, and gassed with 95% O<sub>2</sub> and 5% CO<sub>2</sub>. Tension was recorded isometrically using Ormed UF1 transducers, and displayed on an Ormed Multitrace 4 chart recorder. Tissues were allowed to equilibrate for 90 min before experiments were begun, during which time the resting tension was maintained at 2 g.

# Preparation of haemoglobin

Bovine haemoglobin Type 1 (Sigma) contains a mixture of oxyhaemoglobin and its oxidized derivative methaemoglobin. Pure haemoglobin (oxyhaemoglobin) was prepared by adding to a 1 mM solution of Sigma haemoglobin in distilled water, a 10 fold molar excess of the reducing agent sodium dithionite (Na<sub>2</sub>S<sub>2</sub>O<sub>4</sub>). Sodium dithionite was then removed by dialysis against 100 volumes of distilled water for 2 h at 4°C. The purity of the solutions of haemoglobin was determined spectrophotometrically, and the solutions were frozen in aliquots at -20°C and stored for up to 14 days.

## Measurement of cyclic GMP levels

Endothelium-denuded rings of rabbit aorta were incubated in 50 ml baths containing Krebs solution at 37°C gassed with 95% O<sub>2</sub> and 5% CO<sub>2</sub>. At any time in an experiment these tissues could be removed and plunged immediately into liquid N<sub>2</sub>. Frozen tissues were homogenized in 2 ml ice-cold 6% trichloroacetic acid (TCA), centrifuged at 2,000 g for 10 min at 4°C and the pellet and supernatant separated. TCA was removed from the supernatant by adding 4 ml of 0.5M tri-n-octylamine in freon (1, 1, 2trichlorotrifluoroethane) and vortex mixing until the aqueous phase was pH 5.5-6. The aqueous phase was removed and its cyclic GMP content determined by radioimmunoas-

say using New England Nuclear kits as previously described (Furchgott *et al.*, 1984a). The tissue pellet was resuspended in 1M NaOH, incubated for 30 min at 70°C and protein content determined by the colourimetric method of Bradford (1976).

#### Drugs

Rat synthetic atriopeptin II and phenylephrine were obtained from Sigma and dissolved in double-distilled water. M&B 22,948 (2-0-propoxyphenyl-8-azapurine-6-one) was a generous gift from May and Baker Ltd., and was prepared as a 10<sup>-1</sup>M stock solution in 10% triethanolamine. The final bath concentration of triethanolamine was 0.01% which did not affect tone in control rings.

# Statistical analysis

Results are expressed as the mean  $\pm$  s.e.mean and comparisions were made by means of Student's t test. A probability of 0.05 or less was considered significant.

#### Results

The relaxation of rabbit aortic rings by atriopeptin II and its potentiation by M&B 22,948

Following contraction of endothelium-denuded rings of rabbit aorta with phenylephrine (30–300 nm), exposure to atriopeptin II (30 pM–10nm) induced a dose-dependent relaxation (Figure 1): EC<sub>50</sub> for relaxation was  $8.9 \pm 1.3 \times 10^{-10} \text{M}$  (n=8). The selective cyclic GMP phosphodiesterase inhibitor, M&B 22,948 (100  $\mu$ M), induced a 20.2  $\pm$  1.8% (n=8) relaxation of phenylephrine-contracted endothelium-denuded aortic rings. When tone was re-established to the original level with additional phenylephrine, subsequent relaxation to atriopeptin II (30 pM–10nM) was significantly (P < 0.005) potentiated 2.3 fold (Figure 1): EC<sub>50</sub> for relaxation was  $3.8 \pm 0.6 \times 10^{-10} \text{M}$  (n=8).

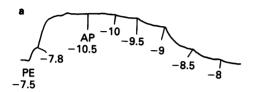
The effect of haemoglobin on atriopeptin II-induced relaxation of rabbit aortic rings

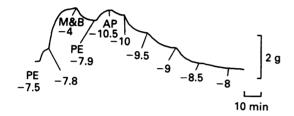
Exposure of phenylephrine (30-300 nM)-contracted rings to haemoglobin  $(10 \,\mu\text{M})$  for  $10-20 \,\text{min}$  produced a  $9.5 \pm 1.7\%$  (n=8) further increase in tone. Following haemoglobin pretreatment atriopeptin II-induced relaxation was not significantly different from that obtained on control rings (Figure 2).

The effect of haemoglobin on the atriopeptin II-induced increase in cyclic GMP content of rabbit aortic rings

Exposure of a ortic rings to a triopeptin II (10 nm) for

3 min induced a 10.9 fold increase in the cyclic GMP content (Table 1). Pretreatment of aortic rings for 20 min with haemoglobin ( $10 \mu M$ ) reduced the resting level of cyclic GMP, but the increase induced by atriopeptin II (10 nM) was unaffected (Table 1).





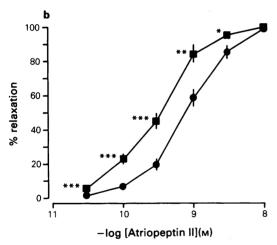


Figure 1 The effect of the selective cyclic GMP phospodiesterase inhibitor, M&B 22,948, on the atriopeptin II-induced relaxation of endothelium-denuded rings of rabbit aorta. (a) Top panel: relaxation of a phenylephrine (PE)-contracted aortic ring by atriopeptin II (AP). Bottom panel: relaxation induced by atriopeptin II following pretreatment with M&B 22,948 (M&B,  $100 \,\mu\text{M}$ ). Cumulative molar concentrations are given in log units. (b) Dose-response curves showing control responses to atriopeptin II ( $\blacksquare$ ) and responses to atriopeptin II following pretreatment for  $10-20 \,\text{min}$  with M&B 22,948 ( $100 \,\mu\text{M}$ ) ( $\blacksquare$ ). Each point is the mean of 8 observations and vertical lines show s.e.mean. \*P < 0.05; \*\*\*P < 0.005.

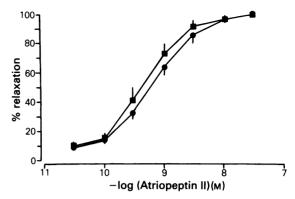


Figure 2 Dose-response curves showing the effects of haemoglobin on the atriopeptin II-induced relaxation of endothelium-denuded rings of rabbit aorta. Control responses to atriopeptin II (●) and responses to atriopeptin II following pretreatment for 10-20 min with haemoglobin (10 μM) (■) are shown. Each point is the mean of 8-10 observations and vertical lines show s.e.mean.

#### Discussion

We found that the relaxation of endothelium-denuded rings of rabbit aorta induced by the atrial natriuretic peptide, atriopeptin II, was potentiated significantly by the selective cyclic GMP phosphodiesterase inhibitor, M&B 22,948. This potentiation was of similar magnitude to the potentiation by M&B 22,948 of relaxation induced by the nitrovasodilators (Kukovetz et al., 1979; Martin et al., 1986a), which also elevate cyclic GMP levels. It suggests a causal role for cyclic GMP in atriopeptin II-induced vasodilatation.

Table 1 Effect of haemoglobin on the atriopeptin II-induced increase in cyclic GMP content of endothelium-denuded rings of rabbit aorta

Stimulus	Pretreatment	Cyclic GMP (pmol g <sup>-1</sup> protein
None (control)	None	$71 \pm 5$ (12)
None `	Нь, 10 μм	$37 \pm 3 ***(10)$
APII, 10 nm	None	$774 \pm 64***(6)$
APII. 10 nm	Hb. 10 им	$770 \pm 76***(6)$

The effects of pretreatment for 20 min with haemoglobin (Hb) were determined on both the basal content of cyclic GMP and on the increased content stimulated by a 3 min exposure to atriopeptin II (APII). The cyclic GMP content is expressed in pmol  $g^{-1}$  protein. Results are expressed as the mean  $\pm$  s.e.mean and the numbers of observations are shown in parentheses. \*\*\*P > 0.001, significantly different from control.

It is not known whether the particulate form of guanylate cyclase has, like the soluble isoenzyme (Craven & De Rubertis, 1978), a ferrous haem-containing receptor site. Methylene blue and cyanide, which inhibit soluble guanylate cyclase, probably by oxidizing or by binding to the haem moiety of the enzyme, respectively (Rapoport & Murad, 1984; Martin et al., 1985), do not inhibit atriopeptin II-induced increases in cyclic GMP content in rat aortic rings (Rapoport et al., 1985). This has been taken as evidence against the presence of haem moieties on particulate guanylate cyclase.

We found that haemoglobin, in concentrations which inhibit smooth muscle relaxation and increases in cyclic GMP induced by EDRF, IF, nitrovasodilators and ultraviolet light (Bowman & Drummond, 1984; Furchgott et al., 1984b; Martin et

al., 1985), had no effect on the atriopeptin II-induced relaxation and associated increases in cyclic GMP in endothelium-denuded rings of rabbit aorta. Our results provide further evidence that a ferrous haem-containing receptor site is not involved in activation of the particulate form of guanylate cyclase.

In conclusion, potentiation by M&B 22,948 is consistent with the view that cyclic GMP mediates the vasodilator action of atriopeptin II. The inability of haemoglobin to block atriopeptin II-induced vasodilatation and associated increases in cyclic GMP in rabbit aorta, suggests that a ferrous haem-containing receptor site is not involved in the activation of particulate guanylate cyclase.

This work was supported by the British Heart Foundation.

### References

- AMBACHE, N., KILLICK, S.W. & ZAR, M.A. (1975). Extraction from ox retractor penis of an inhibitory substance which mimics its atropine-resistant neurogenic relaxation. *Br. J. Pharmac.*, 54, 409-410.
- ARNOLD, W.P., MITTAL, C.K., KATSUKI, S. & MURAD, F. (1977). Nitric oxide activates guanylate cyclase and increases guanosine 3', 5'-cyclic monophosphate levels in various tissue preparations. *Proc. natn. Acad. Sci.*, U.S.A., 74, 3203-3207.
- BOWMAN, A. & DRUMMOND, A.H. (1984). Cyclic GMP mediates neurogenic relaxation in the bovine retractor penis muscle. *Br. J. Pharmac.*, 81, 665-674.
- BOWMAN, A. & GILLESPIE, J.S. (1982). Block of some non-adrenergic inhibitory responses of smooth muscle by a substance from haemolysed erythrocytes. J. Physiol., 328, 11-25.
- BRADFORD, M.M. (1976). A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal. Biochem.*, 72, 248-254.
- BUSSE. R., TROGISCH, G. & BASSENGE, E. (1985). The role of endothelium in the control of vascular tone. *Basic Res. Cardiol.*, 80, 475-490.
- CRAVEN, P.A. & DE RUBERTIS, F.R. (1978). Restoration of the responsiveness of purified guanylate cyclase to nitrosoguanidine, nitric oxide and related activators by heme and hemeproteins. J. biol. Chem., 253, 8466-8443.
- CURRIE, M.G., GELLER, D.M., COLE, B.R., BOYLAN, J.G., YUSHENG, W., HOLMBERG, S.W. & NEEDLEMAN, P. (1983). Bioactive cardiac substances: potent vasorelaxant activity in mammalian atria. *Science*, 221, 71-73.
- FISCUS, R.R., RAPOPORT, R.M., WALDMAN, S.A. & MURAD, F. (1985). Atriopeptin II elevates cyclic GMP, activates cyclic GMP-dependent protein kinase and causes relaxation in rat thoracic aorta. *Biochem. biophys. Acta*, 846, 179-184.
- FURCHGOTT, R.F., CHERRY, P.D., ZAWADZKI, J.V. & JOTH-IANANDAN, D. (1984a). Endothelial cells as mediators of

- vasodilatation of arteries. J. cardiovasc. Pharmac., 6, S336-S343.
- FURCHGOTT, R.F., ERREICH, S.J. & GREENBLATT, E. (1961). The photoactivated relaxation of smooth muscle of rabbit aorta. J. Gen. Physiol., 44, 499-519.
- FURCHGOTT, R.F., MARTIN, W., JOTHIANANDAN, D. & VILLANI, G.M. (1984b). Comparison of endothelium-dependent relaxation by acetylcholine and endothelium-independent relaxation by light in the rabbit aorta. In Proceedings IUPHAR 9th International Congress of Pharmacology. ed. Paton, W., Mitchell, J & Turner, P., Vol. 1, pp. 149-157. Basingstoke Macmillan Press.
- FURCHGOTT, R.F. & ZAWADZKI, J.V. (1980). The obligatory role of endothelial cells in the relaxation of arterial smooth muscle by acetylcholine. *Nature*, **288**, 373-376.
- GILLESPIE, J.S. & MARTIN, W. (1980). A smooth muscle inhibitory material from the bovine retractor penis and rat anococcygeus muscles. J. Physiol., 304, 55-64.
- KARLSSON, J.O.G., AXELSSON, K.L. & ANDERSSON, R.G.G. (1984). Effects of ultraviolet radiation on the tension and the cyclic GMP level of bovine mesenteric arteries. *Life* Sci., 34, 1555-1563.
- KATSUKI, S., ARNOLD, W., MITTAL, C.K. & MURAD, F. (1977). Stimulation of guanylate cyclase by sodium nitroprusside, nitroglycerin and nitric oxide in various tissue preparations and comparision to the effects of sodium azide and hydroxylamine. J. cyclic. Nucleotide Res., 3, 23-25.
- KUKOVETZ, W.R., HOLZMANN, S., WURM, A. & POCH, G. (1979). Evidence for cyclic GMP-mediated relaxant effects of nitro-compounds in coronary smooth muscle. Naunyn-Schmiedebergs Arch. Pharmac., 301, 129-138.
- MARTIN, W., FURCHGOTT, R.F., VILLANI, G.M. & JOTH-IANANDAN, D. (1986a). Phosphodiesterase inhibitors induce endothelium-dependent relaxation of rat and rabbit aorta by potentiating the effects of spontaneously-released endothelium-derived relaxing factor. J. Pharmac. exp. Ther., 237, 539-547.

- MARTIN, W. & SMITH, J.A. (1985). Binding of nitrovasodilators and bovine retractor penis inhibitory factor by haemoglobin and methaemoglobin immobilized on agarose. Br. J. Pharmac. Proc. Suppl., 86, 567P.
- MARTIN, W., SMITH, J.A. & WHITE, D.G. (1986b). The mechanism by which haemoglobin inhibits relaxation of rabbit aorta induced by nitrovasodilators, nitric oxide and bovine retractor penis inhibitory factor. *Br. J. Pharmac.*, 89, 563-571.
- MARTIN, W., VILLANI, G.M., JOTHIANANDAN, D. & FUR-CHGOTT, R.F. (1985). Selective blockade of endothelium-dependent and glyceryl trinitrate-induced relaxation by haemoglobin and by methylene blue in the rabbit aorta. *J. Pharmac. exp. Ther.*, 232, 708-716.
- MITTAL, C.K. (1985). Atriopeptin II and nitrovasodilatormediated shifts in guanosine 3', 5'-cyclic monophosphate in rat thoracic aorta: evidence for involvement of distinct guanylate cyclase pools. Eur. J. Pharmac., 115, 127-128.
- RAPOPORT, R.M., DRAZNIN, M.B. & MURAD, F. (1983).

  Endothelium-dependent vasodilator and nitrovasodilator-induced relaxation may be mediated through cyclic GMP-dependent protein phosphorylation. Trans. Assoc. Am. Physicians, 96, 19-30.
- RAPOPORT, R.M. & MURAD, F. (1983). Agonist-induced endothelium-dependent relaxation in rat thoracic aorta

- may be mediated through cGMP. Circulation Res., 52, 352-357.
- RAPOPORT, R.M. & MURAD, F. (1984). Effect of cyanide on nitrovasodilator-induced relaxation, cyclic GMP accumulation and guanylate cyclase activation in rat aorta. Eur. J. Pharmac., 104, 61-70.
- RAPOPORT, R.M., WALDMAN, S.A., SCHWARTZ, K., WIN-QUIST, R.J. & MURAD, F. (1985). Effects of atrial natriuretic factor, sodium nitroprusside, and acetylcholine on cyclic GMP levels and relaxation in rat aorta. Eur. J. Pharmac., 115, 219-229.
- SCHULTZ, K.D., SCHULTZ, K. & SCHULTZ, G. (1977). Sodium nitroprusside and other smooth muscle relaxants increase cyclic GMP levels in rat ductus deferens. *Nature*, **256**, 750-751.
- WALDMAN, S.A., RAPOPORT, R.M. & MURAD, F. (1984). Atrial natriuretic factor selectively activates particulate guanylate cyclase and elevates cyclic GMP in rat tissues. *J. biol. Chem.*, **259**, 14332–14334.
- WINQUIST, R.J., FAISON, E.P., WALDMAN, S.A., SCH-WARTZ, K., MURAD, F. & RAPOPORT, R.M. (1984). Atrial natriuretic factor elicits an endothelium-independent relaxation and activates particulate guanylate cyclase in vascular smooth muscle. *Proc. natn. Acad. Sci. U.S.A.*, 81, 7661-7664.

Received April 1, 1986. Revised July 2, 1986. Accepted July 23, 1986.)